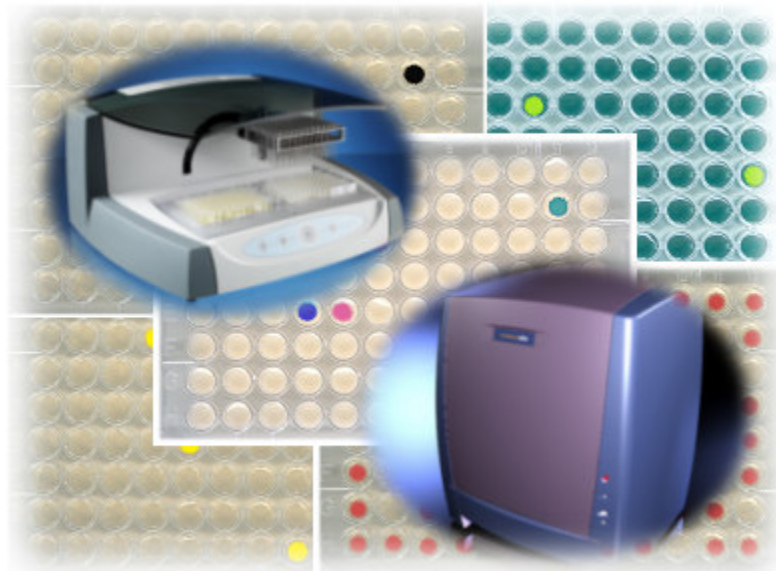


Urine Screening

USERS MANUAL



By Mast Group Ltd.

V1.5 03/06



Contents

Introduction

About This Manual.....	2-5
About The mastascanelite™	3
Daily Operations.....	4
Limitations	5

Chapter 1

mastascan – An Overview	6-9
mastascanelite™ Software	7-9

Chapter 2

Using The mastascanelite™	10-17
Transferring Data To The mastascanelite™	10
Import Data Using Import/Export.....	10-12
Input Data Using Patient Data.....	12-17

Chapter 3

The Read Program	18-22
Urine Screening Read Overview	18
The Reading Process	18-22

Chapter 4

The Generate ID Program	23-26
Identifications overview	23
Generating identifications.....	24-26

Chapter 5

Reporting Results	26-29
Export Using Impex	27-29
Printing Results Using Reports	30

Chapter 6

Trouble Shooting	31-32
Problems And Solutions	31
Contacting Mast	32



About This Manual

This manual is a step by step guide on how to routinely operate the **mastascanelite™**. To help make this guide easy to follow key words and phrases have been displayed in different colours/fonts. Below is a list of examples

Example	Description
OK, Cancel, Add Threshold	Bold/blue are used for the names of command buttons
<i>Textbox, DataField</i>	Italics are used for references made to a box in which you enter data.
Urines_1, Streptococci	Bold/Red type is used for words the user is instructed to type.
[Tab]. [Return]	Square brackets are used to indicate keys on the keyboard.



About The **mastascanelite™**

mastascanelite™ (Fig. 1) is a sophisticated and flexible computerised image analysis system capable of reading, interpreting and reporting antibiotic susceptibility and bacterial identification carried out using multipoint technology.

Urinary Tract Infection (UTI) is defined as the presence of significant bacteruria, and urine specimens may comprise the majority of the routine laboratory's samples. The **mastascanelite™** Urine Screen System provides a rapid and accurate method of screening urine specimens for the presence of bacteria, and providing an identification and antibiotic susceptibility test on the organisms present in the samples.



Figure 1. The **mastascanelite™**

The **mastascanelite™** significantly reduces reading and interpretation errors and removes the possibility of transcription inaccuracies. The system substantially reduces the time required each day to read, interpret and report the day's urine samples, whilst still leaving you in full control of each stage. You have the facility to override any computer-generated interpretation or measurement.

The system is operated by an powerful IBM compatible PC using Microsoft Windows as the operating system. The software is especially designed to provide a user friendly Graphical Interface that can be driven by both a mouse and keyboard combination.



Introduction

Daily Operation

On a daily basis patient and specimen details are entered onto a template. Alternatively, **mastascanelite™** can be interfaced to laboratory computer systems via a bi-directional link where patient details can be automatically transferred to the **mastascanelite™**. Once the set of urine screen plates has been read, the results are relayed back to the laboratory computer for reporting. Each urine sample is treated as a separate record and saved within the mastascanelite database.

Each urine specimen is inoculated into a well in a series of microtitre plates using an automated multipoint inoculator e.g., The **multipointelite™** (Product Code SCAN4000). The plates are then incubated for 18-24 hours at 37°C.

Each urine screen plate is placed in the motorised specimen drawer. The **mastascanelite™** reads each plate in order, and interprets the sensitivity to each antibiotic by reference to a non-selective control plate, and the identifications by colour reactions. The urine screen system incorporates a manual interpretation stage, allowing total user confidence in detection of mixed cultures and contamination etc. Total control of the interpretations is maintained by the override facility. User definable observations are also available for attachment to individual samples.

The **mastascanelite™** incorporates a powerful user definable reporting facility. Results are stored in the database used by the system which can either be exported back to the Host Laboratory System or displayed and/or printed a various formats using a standard Windows based printer



Limitations

Any antibiotic combination can be used with **mastascanelite™**. The system allows antibiotic plates to be read in any pre-defined order.

The agar in the microtitre plates must be of equal depth, both within the same plate, and also between the different plates. The agar must also not contain any bubbles or impurities and be free from bacterial/fungal contamination. Presence of these and differences in agar depth will result in inaccurate results being obtained. It is for this reason that Mast Group Ltd expressly recommends the use of Mast Rediprep Urine Screening plates.



Chapter 1

mastascanelite™ elite Overview

PRECAUTIONS

The mastascanelite software is specially designed to look and feel like a standard Windows© software package. It allows user-friendly operation by all laboratory staff irrespective of the level of computer literacy. This chapter provides a general outline to operating the mastascanelite software.

The **mastascanelite™** software consists of a suite of applications. This is a group of related programs that control data flow and data acquisition.

Caution

Contact Mast Group Ltd for advice on software that can be installed. Although it is unlikely that installation of none "Mast" software will create any operation problems it is recommended that the software is described to Mast Group Ltd to ensure compatibility.

The End User is responsible for ensuring that any required End User Licences are in place for software installed on the system that was not provided by Mast Group Ltd.



Chapter 1

MASTASCANELITE™ SOFTWARE

When switching on the power to the instrument, it is important to switch the **mastascanelite™** on before turning on the computer. The power switch on the **mastascanelite™** is located on the rear of the instrument, adjacent to the power supply cable.

When the computer has loaded Windows©, locate the **Command Centre** (figure 1.1) icon on the screen.



Figure 1.1. The Command Centre Icon

Double click on this icon to open the **mastascanelite™** log-on screen (figure 1.2)

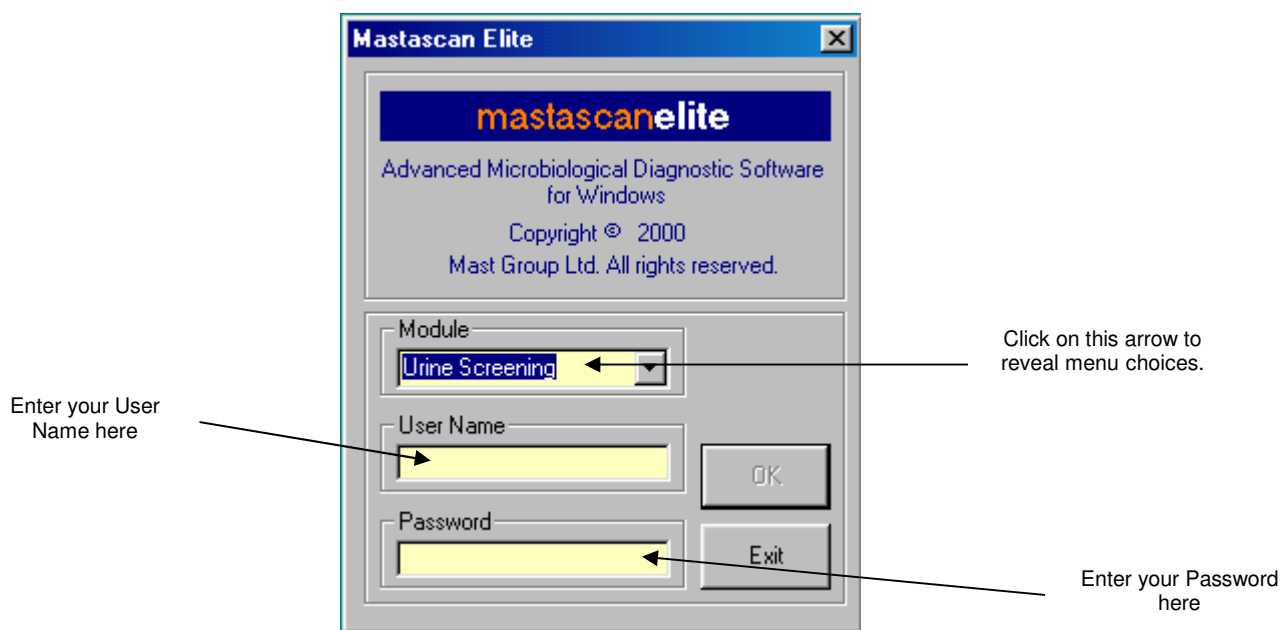


Figure 1.2. The **mastascanelite™** log-on screen.

If you have purchased other modules, select Urine Screening from the menu which appears when you click on the down pointing arrow. Click in the User Name field, and type in the user name that you have been given. If you do not have a user name then contact your Instrument Supervisor and click on the Exit button to unload the software.



Chapter 1

Press the [Tab] key. Then enter the Password with which you have been supplied. Press the [Enter] key, or click on **OK**. This will then display the **mastascanelite™ Command Centre** (figure 1.3)

N.B. The **mastascanelite™** administrator will liaise with staff regarding user names, passwords and access levels.

The software is divided up into individual programs, each represented by a button on the toolbar. You enter these programs by clicking on the appropriate button. The number of buttons displayed will vary dependent upon the level of access the user has been given.



Figure 1.3. The **mastascanelite™** elite Command Centre.

If you are unsure what each button will launch then hold the cursor over the button and after a second a label will appear, known as a tool tip, displaying the program name.

Dependent upon the specified level of access, the following icons may be visible.

Administration (figure 1.4) is where user defaults and options are set up.



Figure 1.4



Chapter 1

Configuration (figure 1.5) is where the data for the **mastascanelite™** is entered and edited. Information such as Antibiotic codes, Organism codes, Antibiotic and Identification working lists is stored here.



Figure 1.5

Identification Database (figure 1.6) is where the identification probabilities are stored and edited.



Figure 1.6

For more information on these three functions, please refer to the Set-up and Calibration manual.

Patient Data (figure 1.7) is the function for manual entry of patient records, and for editing automatically entered data.



Figure 1.7

Import & Export (figure 1.8) This function is the interface. It allows transfer of data between the **mastascanelite™** and the host computer system.



Figure 1.8

Read (figure 1.9) is the program that actually reads the plates, and interprets the results.



Figure 1.9

Generate Identifications (figure 1.10) enables the **mastascanelite™** to interpret the results from the identification tests, and produce an actual identification for a bacterial sample.



Figure 1.10

Data Archiver transfer plate runs and all associated data from the mast database and stores them in an archive, enabling the instrument to retain optimal performance.



Figure 1.11

Chapter 2

Using the mastascanelite™

TRANSFERRING DATA TO THE mastascanelite™

Turn on the mastascanelite™ and computer, and open up the **Command Centre** as previously mentioned. The first process is to enter the patient data into the mastascanelite™. This can be done in two ways, either by using the manual **Patient Data** program, or by using the automated **Import & Export** program.

IMPORTING DATA USING IMPORT/EXPORT

From the Command Centre click on the **Impex** Button. The **Import/Export** program will be loaded (figure 2.1)



Figure 2.1. The import & Export screen

Click on **Import** from the Import/Export Form. The Import screen will appear (figure 3.2)

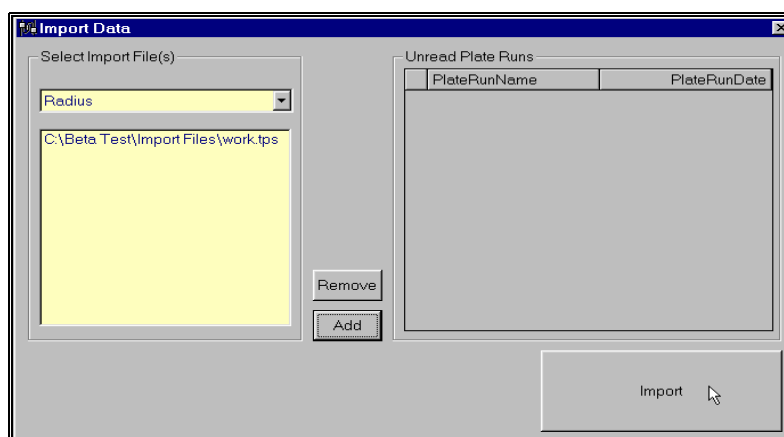


Figure 2.2. The Import screen



Chapter 2

The Import form requires the following information to import a file:

- The plate runs view
- The file format to import to
- The file(s) to import.

Viewing Plate Runs

The plate runs view on the right hand side of the form allows you to check to see which plate runs have been imported. As it displays all plate runs, it will also show current unread plate runs.

Select A File Format

To select a file format, click on the arrow on the drop down list and select the desired file format.

Caution

Be careful when selecting the file format. If the format is incorrect then the imported data is also likely to be incorrect.

It is critical that the correct format is selected. This can be set in the import options form (See below)

Select File(s) to Import

1. **mastascanelite™** will import any number of files in one batch.
2. Click on the **Add** on the import window (figure 2.2).
3. The open dialogue box will appear (figure2.3)
4. Select a file from the list.
5. Click on the **Open** Button.
6. The file will be added to the list.
7. Repeat until all desired files have been added.

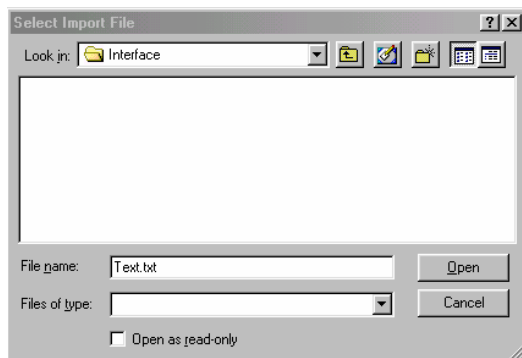


Figure 2.3 Select Import File



Chapter 2

Importing Data

1. When all the above options are completed, the selected files can be imported.
2. Click on the **Import** Button.
3. The data will automatically be imported into the **mastascanelite™** database.

INPUT DATA USING PATIENT DATA

Patient Data Overview

The **Patient Data** Entry facility is the hub of the **mastascanelite™** system. Patient and specimen details are entered on a template structure and stored in the database as batches of patients, known as plate runs.

Using the **mastascanelite™** each patient or specimen is treated as a separate record. Attached to each record is the essential configuration data enabling **mastascanelite™** to read and interpret each sensitivity plate irrespective of the type of media, organism or antibiotic configuration.

The Patient Data Program is used by the **mastascanelite™** to read the corresponding set of antibiotic and identification plates, resulting sensitivities and identifications are linked to the patient and the specimen. Patient and specimen details together with the interpretations may be viewed and modified at any time.

Before we continue, a brief description of the structure and methods behind the patient data module have been included below to help in understanding the system.

The Patient Data Model

The Patient Data Model consists of the following main elements of data:

- | | | |
|------------|---|--|
| Plate Runs | - | a grouping system for batches of patients, each with an isolate to be tested against a set of antibiotic/identification tests |
| Plate Sets | - | each set is a group of antibiotic and identification tests to be used against each isolate |
| Patients | - | all the patient information is stored on the mastascanelite™ elite including: name, lab/hospital number, gender, date of birth etc. |
| Isolates | - | Each isolate is one urine sample allocated to a patient |

These elements are linked together in a hierarchical way so that: –

- 1 Plate Run contains up to 96 patients with 1 Plate Set assigned to it.
- 1 Patient contains 1 or more Isolates (Specimens)



Chapter 2

Loading the Patient Data Module.

1. Click on the *Patient Data* button (figure 1.7) in the toolbar of the **Command Centre**. (If you are unsure which button loads the **Patient Data** module, move the mouse cursor over each button for a few seconds and a label will appear with the name of the module).
2. The main **Patient Data** window (figure 2.4) will appear.

Figure 2.4. The main Patient Data window

Adding a Plate Run

1. From the **Patient Data** main windows (figure 2.4)
2. Click on the window "1 Plate Runs". The Plate Run window (figure 2.5) will appear in front.
3. Press the [CTRL]+[A] keys or click on the **Add** button on the Plate Run Form.
4. The Add Plate Run form will appear (figure 2.6).

Chapter 2

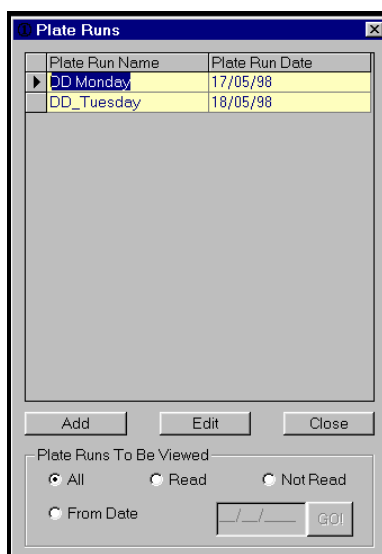
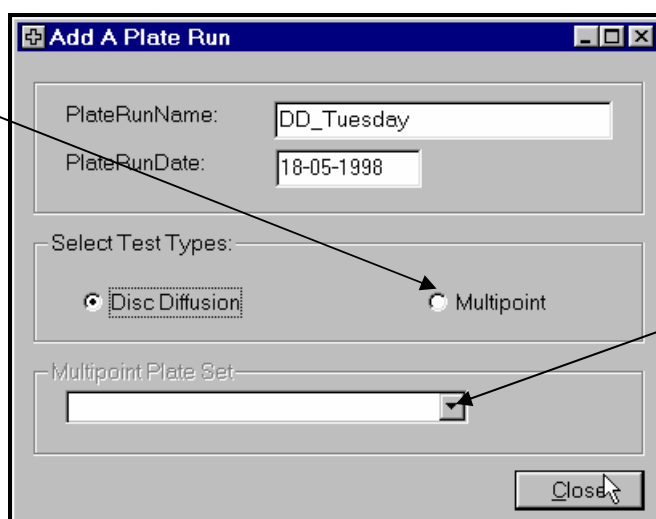


Figure 2.5. The Plate Run window

Click this check box to select multipoint



Click on here to reveal the Plate Sets

Figure 2.6. The Add Plate Run window.

5. In the *Plate Run Name* field, type the name of the new plate run.
6. Click in the *Plate Run Date* field, which automatically inserts the current date.
7. Click on the *Multipoint* check field (This will only be present if other modules are installed)
8. Select the correct Plate Set by clicking on the down arrow to reveal the menu.
9. Click on the **Close** Button.
10. The Plate Run will be added to the database
11. Repeat steps 3 -10 for all the Plate Runs to be entered.



Chapter 2

Editing Plate Run Data

1. On the Plate Run window (figure 2.5) click on the **Edit** button.
2. You can now edit any of the Plate Run Names in the Plate Run List.
3. Click on the same button, now called **Finish Edit** when you have finished.

Entering Patient data

The second window in the patient data module is the template structure in which individual patient details can be viewed and modified on. The Patient Data Entry Template (figure 2.7) comprises of a number of pre-defined fields that correspond to specific patient data.

To open this window double click on the plate run in the plate run window (figure 2.3), select Patient then View Patient Data from the menu at the top of the main patient data environment screen, or click on the grey bar "Patient Data"

Click here to display a list of Plate Runs

Patient Data

Plate Run Details
Select Plate Run: 6 Date: 06/02/02 **Read**

Patient Details

1
2
3
4
5
6
7
8
9
10
11

Lab Number: 4
Hospital Number:
Surname:
Forename:
DOB:

Gender
☐ Male
☐ Female
☒ Unknown

Add Patient Auto Fill
Edit Patient Auto Isolates

Isolate Details

UNKNOWN

	Antibiotic Name	Antibiotic Code	Dilution	Result
▶	Ampicillin	AP	32	#
	Augmentin	AUG	32	#
	CEFTRIAXONE	CRO	32	#
	CEFUROXIME	CXM	32	#
	CEPHALEXIN	CFX	32	#
	Ciprofloxacin	CIP	8	#
	Ciprofloxacin	CIP	1	#
	GENTAMICIN	GM	8	#
	GENTAMICIN	GM	1	#

Add Isolate
Change Isolate
Edit Zone
Edit Antibiotic
Edit Plate Set

UNKNOWN UNKNOWN UNKNOWN

Figure 3.6. The Patient Data window.

Chapter 2

1. By clicking on the down arrow, a list of current Plate Runs will be displayed.
2. Click on the Plate Run you wish to populate with data.
3. Click on the **Auto Fill** button. This will open up the *Auto Fill* window (figure 2.7)

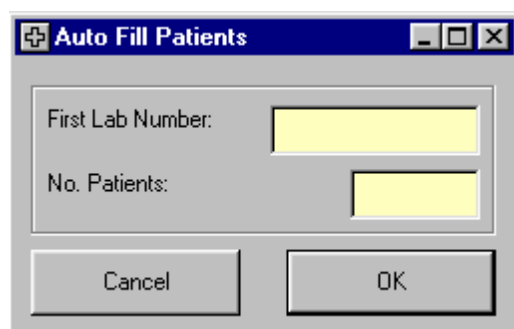


Figure 2.7 Auto Fill Window

4. Enter the first Laboratory Number into the *First Lab Number* text box, the number may contain alphabetical characters and numbers.
5. Enter the number of consecutive patient samples into the *Number of Patients* text box. Individual samples may also be added in this way.
6. Repeat this until all patient samples have been entered, to a maximum of 96.
7. Click on the **Auto Isolate** button. A message will appear on the screen (figure 2.8)

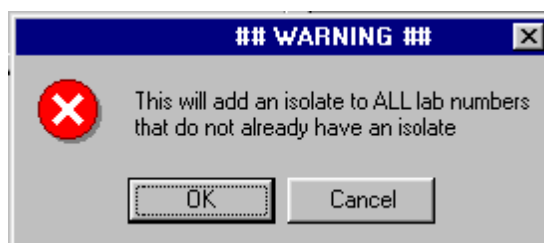


Figure 2.8 Auto Isolate window

8. Click on the **OK** button. The software will now scroll through the patients adding the set of antibiotic and identifications as contained within the Plate Set.

Now, if you click on one of the patients in the left window, the range of tests will appear in the bottom window.

Modifying Patient Details

As we mentioned earlier, both automatically imported, and manually created data can be edited through the Patient Data module:

1. After selecting the appropriate Plate Run in the Plate Run window.



2. Click on the reference number of the patient whose details you wish to modify from the list box on the left hand side of the window.
3. Click on the *Edit Patient* Button.

Chapter 2

4. The fields displaying the patient information will now be live (figure 2.9).
5. You can change any part of the patient information except the Lab Number.
6. When all modifications have been made click on the *Finish Edit* button.
7. All changes will have been recorded and can now be viewed as normal.

The screenshot shows a software window titled "Patient Data". It contains two main sections: "Plate Run Details" and "Patient Details".

Plate Run Details:

- Select Plate Run: DD Monday (dropdown menu)
- Date: 17/05/98 (text field)
- Not Read (button)

Patient Details:

- Left List Box:** Contains two entries, "mst0001/01" (highlighted) and "mst0001/02".
- Form Fields:**
 - Lab Number: mst0001/01 (text field)
 - Hospital Number: L20 (text field)
 - Surname: Kind (text field)
 - Forename: David (text field)
 - DOB: 07-Sep-72 (text field)
 - Admitted to emergency ward with infected upper arm. (text area)
- Gender Section:**
 - Male (selected radio button)
 - Female (radio button)
 - Unknown (radio button)
- Buttons:** "Add Patient" and "Finish Edit" (both disabled).

Figure 2.9. The Edit Patient window.

Once the Plate Run has the contents and structure you require, click on the **X** at the top left corner of the screen to close the program. The instrument is now ready to read.



Chapter 3

The Read Program

URINE SCREENING READ OVERVIEW

The **mastascanelite™** can be configured with any interpretative or test criteria that the laboratory desires when reading plates as described in the Configuration Manual.

The **mastascanelite™** comprises of a specialised state of the art image analysis system. Each test plate is positioned in the motorised specimen drawer in turn. The camera relays the image of the sensitivity test plate to the advanced processing system for analysis.

The image is adjusted automatically for each plate and along with the specialised lighting system enables the **mastascanelite™** to read the plate. As discussed earlier, the Plate Set contains the essential configuration that enables **mastascanelite™** elite to read and interpret a urine screening plate.

The mouse controls the reading of Multipoint plates along with certain short cut keys. Once **mastascanelite™** has read each plate the computer generated results are overlaid on the image of the Multipoint plate. The user maintains full control during the reading and interpretation of each Pin Position by use of the override and observations options.

THE READING PROCESS

Opening the Read Program

From the Command Centre click on the **Read** button on the toolbar.

The Multipoint reading module window will appear with a list of unread plate runs in the plate run screen (figure 3.1)

The Plate Runs screen is similar to that in the patient data module, except at the reading program stage you cannot alter any details to plate runs.

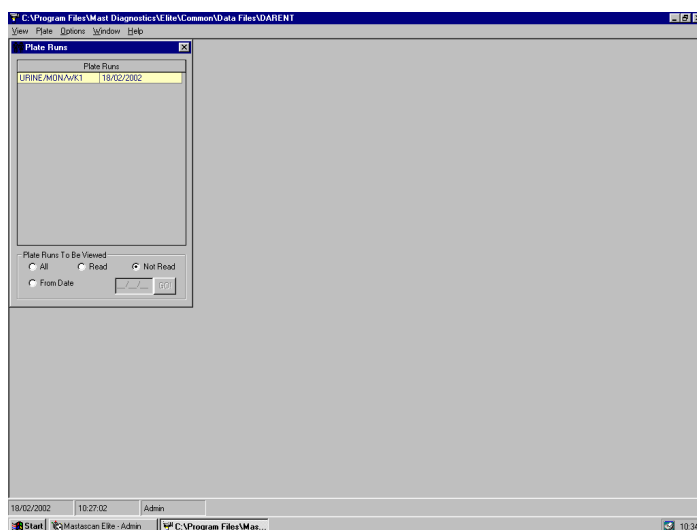


Figure 3.1. Urine Read Window



Chapter 3

Reading a Plate Run

To read a plate run, double click on the desired plate run . The **mastascanelite™** will then prompt you to read either breakpoints or identifications (figure 3.2)

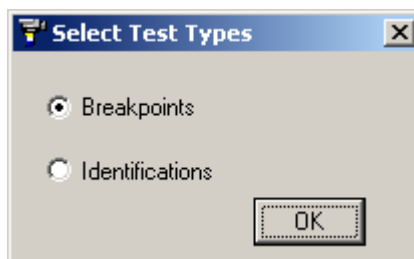


Figure 3.2 Identification/Breakpoint Choice

For urine screening it is recommended that the breakpoints are read first, this will screen out all negative urine samples before reading the remaining plates:

Reading Options

The keyboard can be used to operate the read program:

- [F5] – instructs the machine to open the drawer and go back a plate
- [F6] – read the current plate in the machine
- [F7] – open the drawer and go to the next plate

The mouse can operate the read program:

- Click on the **Left Arrow** which has the same function as [F5]
- Click on the **Camera** button to read the current plate in the machine
- Click on the **Right Arrow** to proceed to the next plate.

On the **mastascanelite™** elite box there are three buttons that can be used to operate the machine:

- Press  to open or close the drawer.
- Press  to read the current plate .
- Press  to move onto the next plate to be read



Chapter 3

Reading Breakpoints

The following instructions refer to using the mouse for all operations but the keyboard and **mastascanelite™** elite buttons can be used as mentioned previously.

It is important to ensure that when using the system for Urine Screening the lightblocker template is attached to the drawer mechanism, this must be in place at all times when reading both antibiotic and id plates.

Check to make sure the drawer is in the correct position, slide the template until it is aligned with the 12cm marker.

Click on **Breakpoints** (figure 3.2) the drawer will open and the main Read window (figure 3.3) will appear displaying the pin positions for all the wells.

Ensure the infra red black background plate is positioned in the drawer before reading.

Place the background plate in the machine so that it rests in the drawer in the holder for the plates.

Click the **Camera** icon to read the plate The machine will then read the plate.

Click on the **Right Arrow** to move to the next plate. Replace the plate with the next which is the growth control plate.

Click on the **Camera** icon to close the drawer and read the plate.

Chapter 3

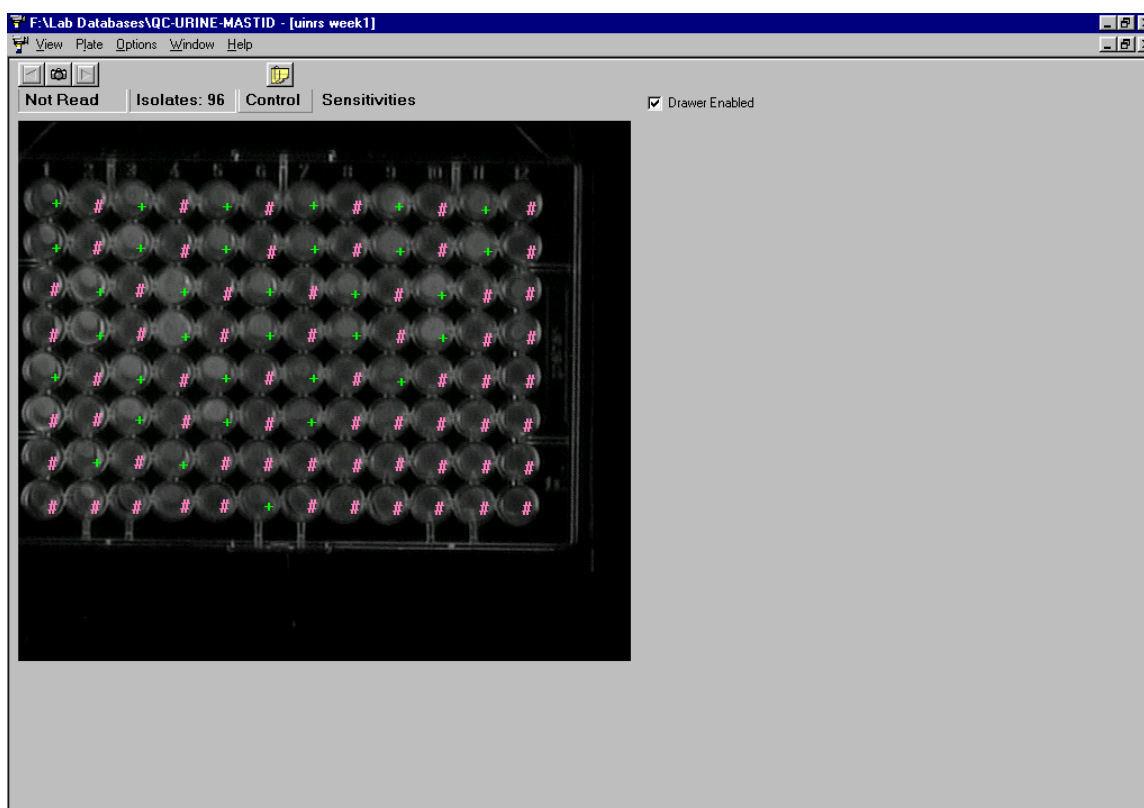


Figure 3.3. Main Reading window (reading the growth control plate)

The **mastascanelite™ elite** will display all growth on wells as “+” and all no growth as “#”.

If any result is incorrect then double click on the result to change.

Click on the **Right Arrow** button to go to the next plate which will usually be the CLED plate.

Click on the **Camera** icon and a message will state the plate is read manually. Visually check the plate and add CLED observations to the relevant wells by using a right click on the well then left click the required observation. **If the patient information was downloaded from the HOST system then if transmitted, the white/red/epithelial cell count is displayed if the mouse is moved onto a well. This feature allows the technician to decide the clinical significance of any growth on a well.**

Click on the **Right Arrow** button to move to the next plate which will be the first antibiotic plate.

Insert the relevant plate and click on the **Camera** button to read the plate. Note at this point when the machine reads, any wells that were negative on the control plate will be ignored from this point onwards. After the plate has been read any



incorrect results may be altered by double clicking on the well. Any wells that were changed from negative to positive on the control plate will always be displayed as “?” which refers to a manual result must be entered by double clicking on the “?” to change to positive or a further double click to change to negative.

Chapter 3

Click on the **Right Arrow** button to go to the next plate and repeat the whole reading process for the remaining antibiotic plates. After the last plate has been read if the **Right Arrow** is clicked again then a message will pop up “Last Plate”. Click **OK** to close the message. Click on the **X** of the inner Read window to close.

Note: When using a plate set containing multiple dilutions of the same antibiotic, if a sensitive result occurs at a higher dilution and a resistant result occurs at a lower dilution a message will pop up, “A resistant result has occurred after a sensitive result do you want to stop and correct?”. Click on **No** to ignore and proceed or click on **Yes** to stop and correct. If **Yes** is selected a list of disputed results will be displayed. Select the appropriate results and correct them.

Once all antibiotic plates have been read the **mastascanelite™** can be used to read and interpret the identification plates.

Reading Antibiotic Plates

Double click on the same plate run name that has been read for antibiotics in the Read window (figure 3.1). A message will pop up asking whether identifications or breakpoints are to be read (figure 3.2). Select the identifications option.

The drawer will open. Take out the black background and replace it with the light grey background.

Place the first identification plate to be read on the drawer. Note, near the top of the reading screen the name of the plate to be read is visible to ensure the correct plate is put into the machine.

Click on the **Camera** icon to read the plate. The identification plates are read in colour. The colour of the reactions may not always be the same colour as when read by eye. The machine is calibrated to give the biggest colour difference between positive and negative reactions so alternative colours may have been used.

Chapter 3

Figure 3.4 shows an identification plate that has been read.

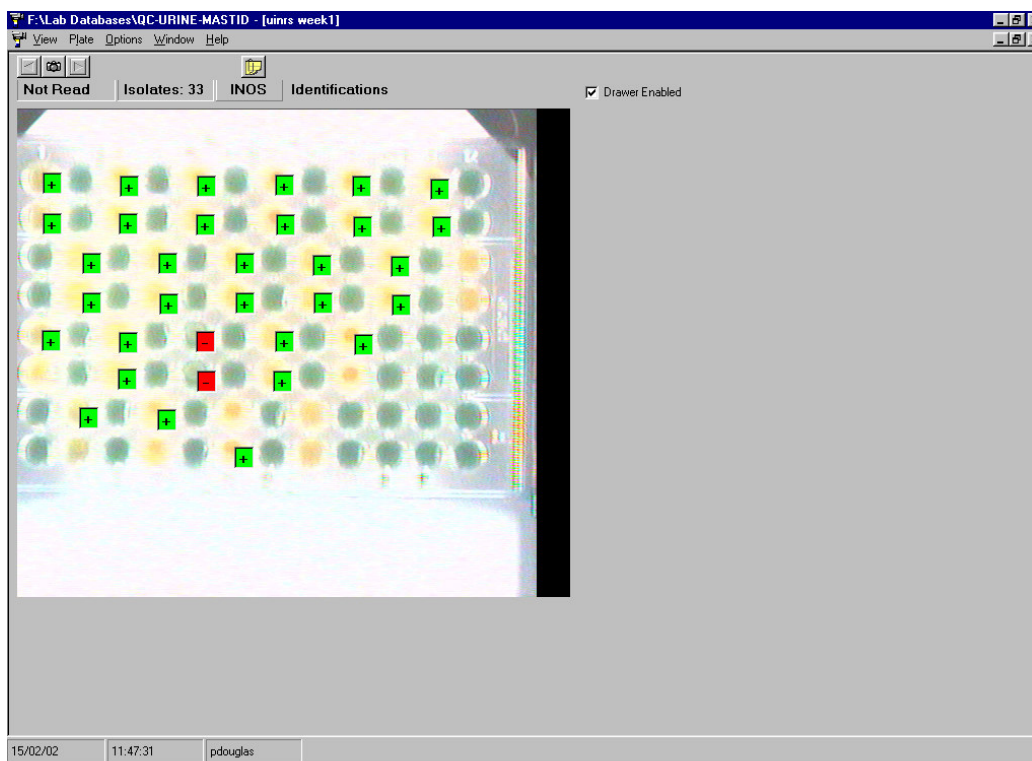


Figure 3.4. Reading An Identification Plate

If any of the results are incorrect double click on them to change the result. If any of the results are displayed as, "#", the **mastascanelite™** has not been able to differentiate whether the result should be positive or negative and manual input is required, double click to change the result to positive or double click again to change to negative.

Read all the identification plates in the same way.

When reading is complete click on **X** of the main Read window to close down the Read Program.



The Generate ID Program

IDENTIFICATIONS OVERVIEW

Identification Database

The **mastascanelite™** contains a database that stores all the probabilities for each colour reaction against known strains of bacteria using the Mast pre-poured media plates. The database can be modified to suit the needs of the customer e.g. additional tests or known strains.

Using the results of each colour reaction in a plate run the Generate ID program can interpret the data to produce a list of bacterial identifications against laboratory number and display the percentage likelihood e.g.:

<i>Identification</i>	<i>Likelihood</i>	<i>Comment</i>
<i>Escherichia coli</i>	95.69%	Excellent Identification

Once the identifications have been generated the results can be viewed and conflicting results that reduce the likelihood of an identification can be displayed and altered if necessary, e.g.

<i>Escherichia coli</i>	38.64%	Poor ID	ONPG –
<i>Escherichia coli</i>	62.52%	Good ID	ONPG changed to +

Reporting Results

Once all the identifications have been generated the **mastascanelite™** can report the results in several ways:

- The results can be exported to the laboratory host system.
- The **mastascanelite™** can display the results on screen.
- The results can be printed using the built in report facility on the desktop of the PC



Chapter 4

GENERATING IDENTIFICATIONS

Starting from the **Command Centre** click on the icon for **Generate ID's** (figure 1.10)

The Generate ID window will open (figure 4.1)

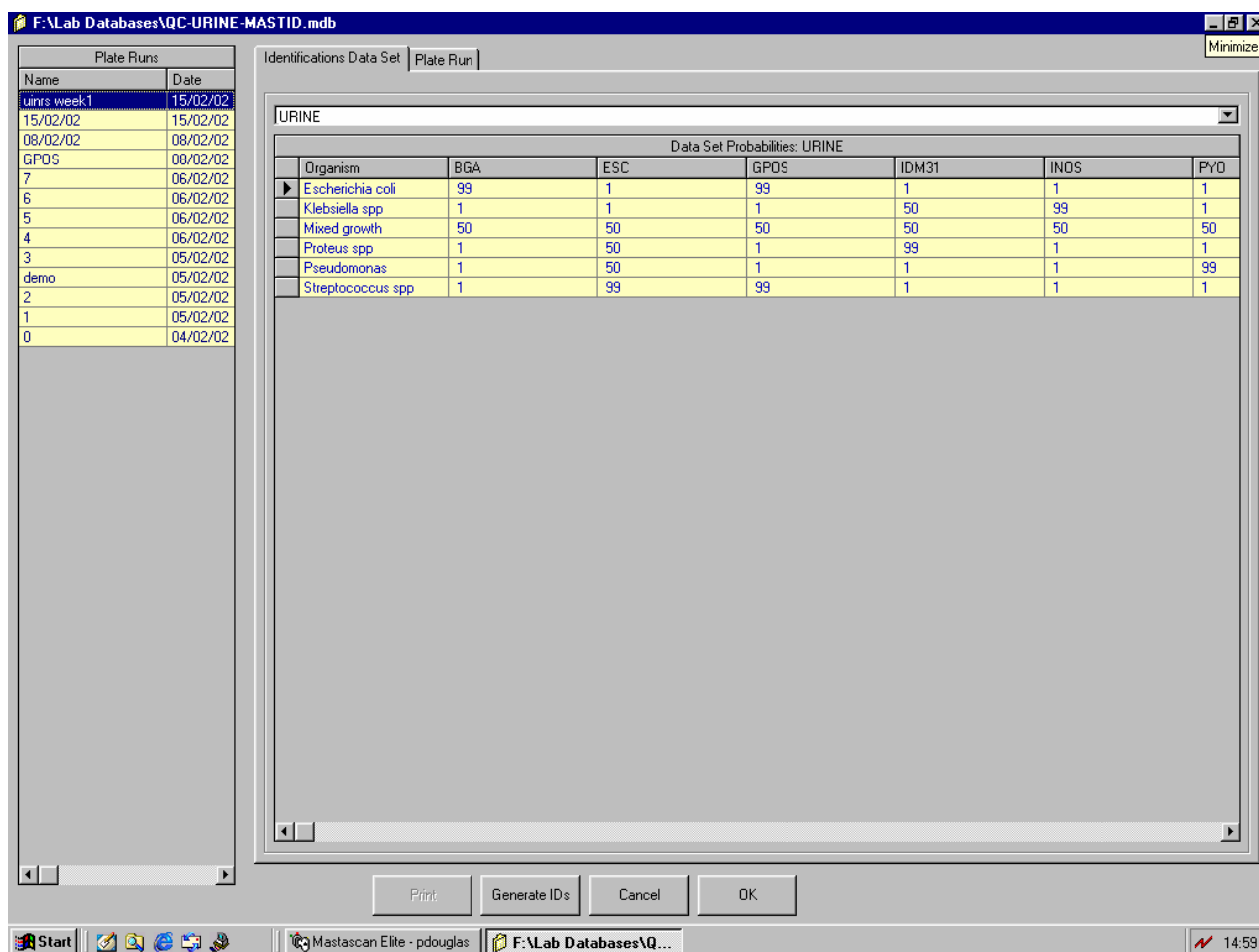


Figure 4.1. Generate ID window

Highlight the Plate Run to be used.

From the drop down menu select the ID Data Set. As standard the **mastascanelite™** contains two Data Sets; Urine and MASTID15. Customer specific Data Sets have to be created if these are not to be used, as detailed in the Urine Screening Configuration Manual.

Click on the **Generate ID's** button. Within a few seconds all the samples that had growth in the Control Plate will have an identification attached to them. To view the results click on the **Plate Run** tab near the top of the window and they will be displayed in a list going downwards (figure 4.2).



Chapter 4

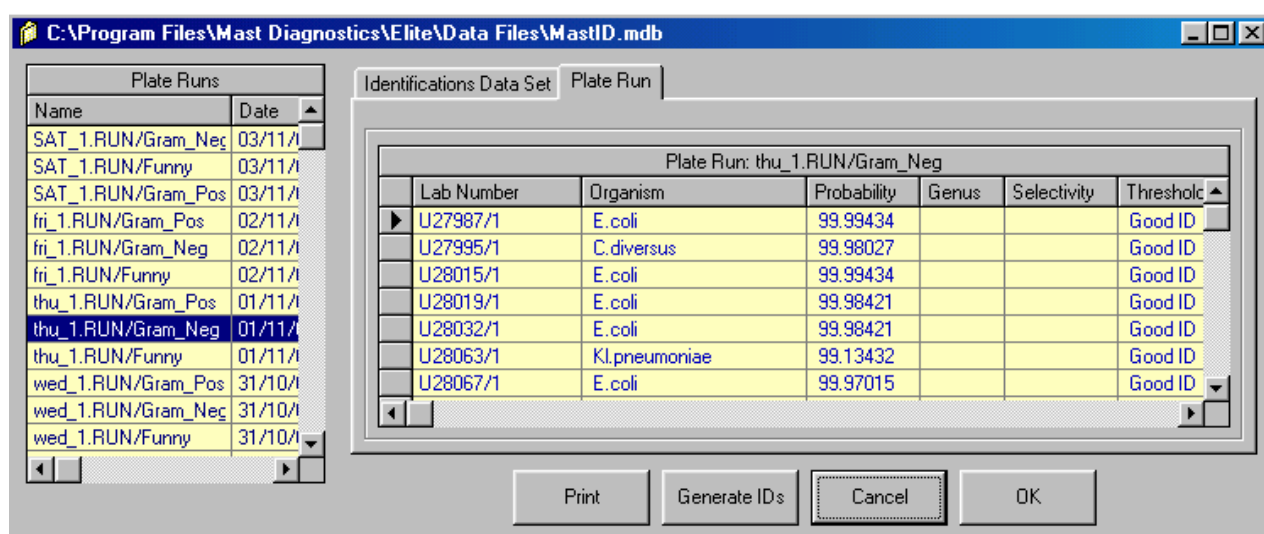


Figure 4.2. Generated Identifications

If an interpretation of one of more results needs to be queried double click on the laboratory number to be checked.

A second window will appear displaying all the results for the specific laboratory number (figure 4.3)

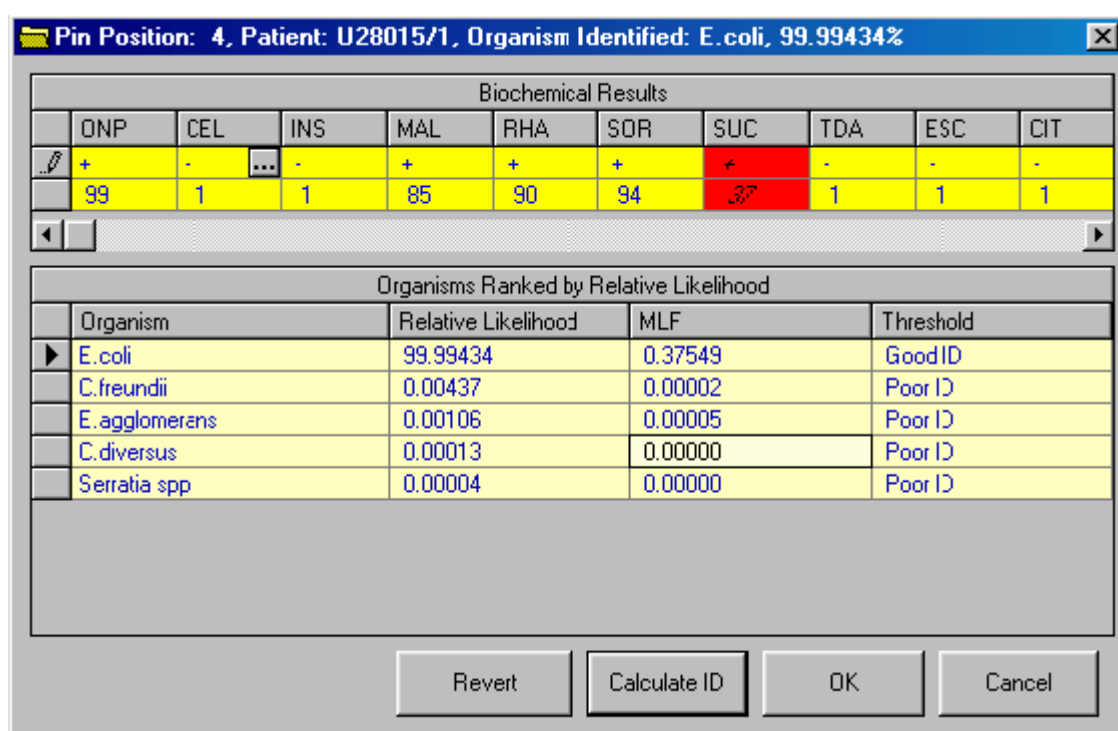


Figure 4.3. Detailed Interpretation of a Single Laboratory Number

All results displayed in yellow are the correct interpretation for the species identified. The five most likely identifications are listed with the most likely at the top.



Chapter 4

Results displayed in red are those that are aberrant for the identification displayed. To alter a result click on the box with the result in and click on the button [...]. After the results has changed click on the **Calculate ID** button to see the outcome of the changes.

To accept the changes click **OK**. To cancel the changes click on **Revert**.

Click **OK** to close the window.

To save the identifications click **OK** in the main window (figure 4.2). If you do not wish to save the interpretations, click on **Cancel**

Reporting the Results

Below are the details of all the methods that can be used by the **mastascanelite™** to report the results.

EXPORT USING IMPEX

From the Command Centre click on **IMPEX** button (figure 1.8) to load the program. When the Import/Export window pops up click on **Export** to bring the next window up. (figure5.1)

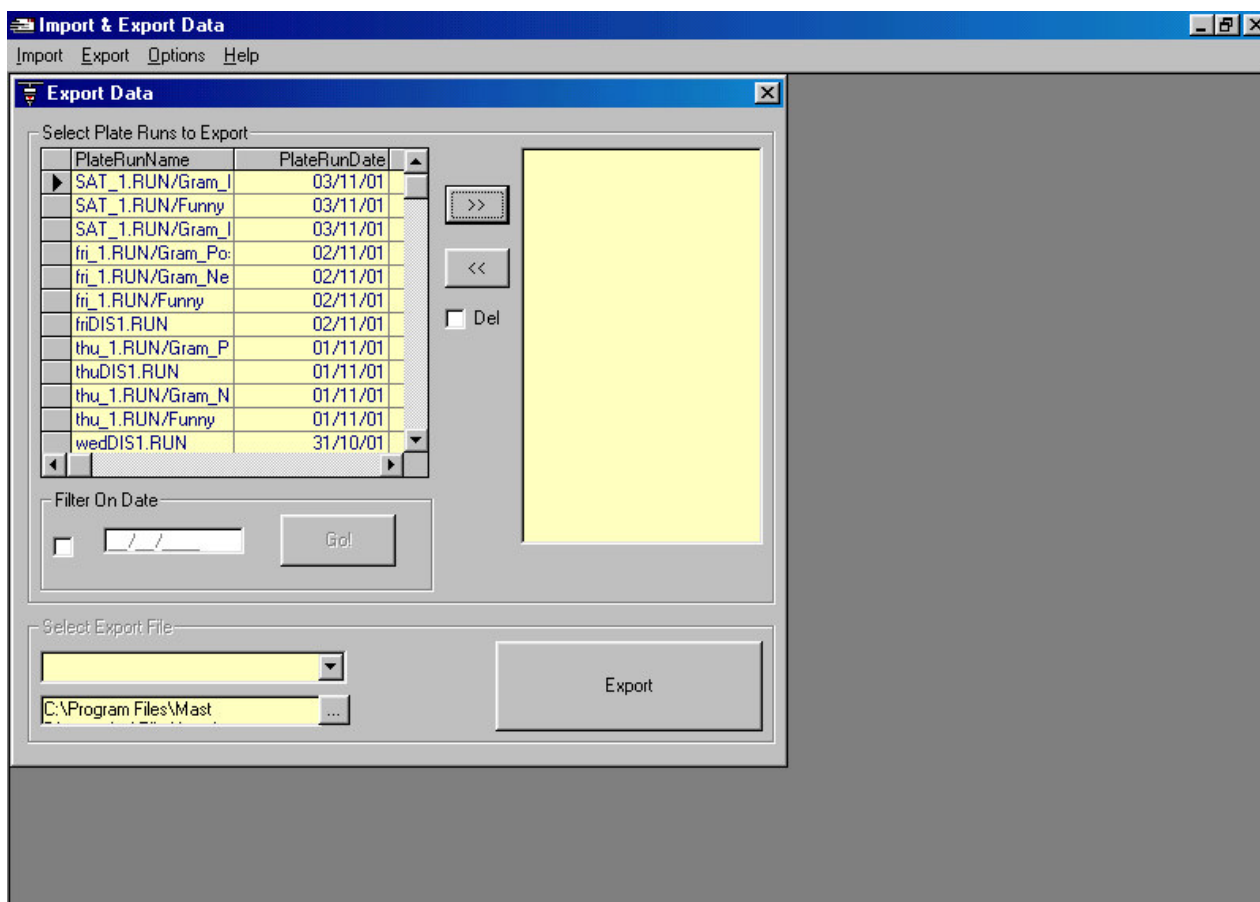


Figure 5.1. Export Window

Select a plate run to be exported then click [**>>**]. Repeat this for all the plate runs to be exported. This will prepare all the plate runs required for transfer to the Laboratory Host System.

There is an option to filter the list of plate runs by date before preparing for transfer. To do this type in a date e.g. **08/09/2002** in the date field below the list of plate runs the press **Go!**. This will now list only the plate runs that were created on the date entered in the filter.



Chapter 5

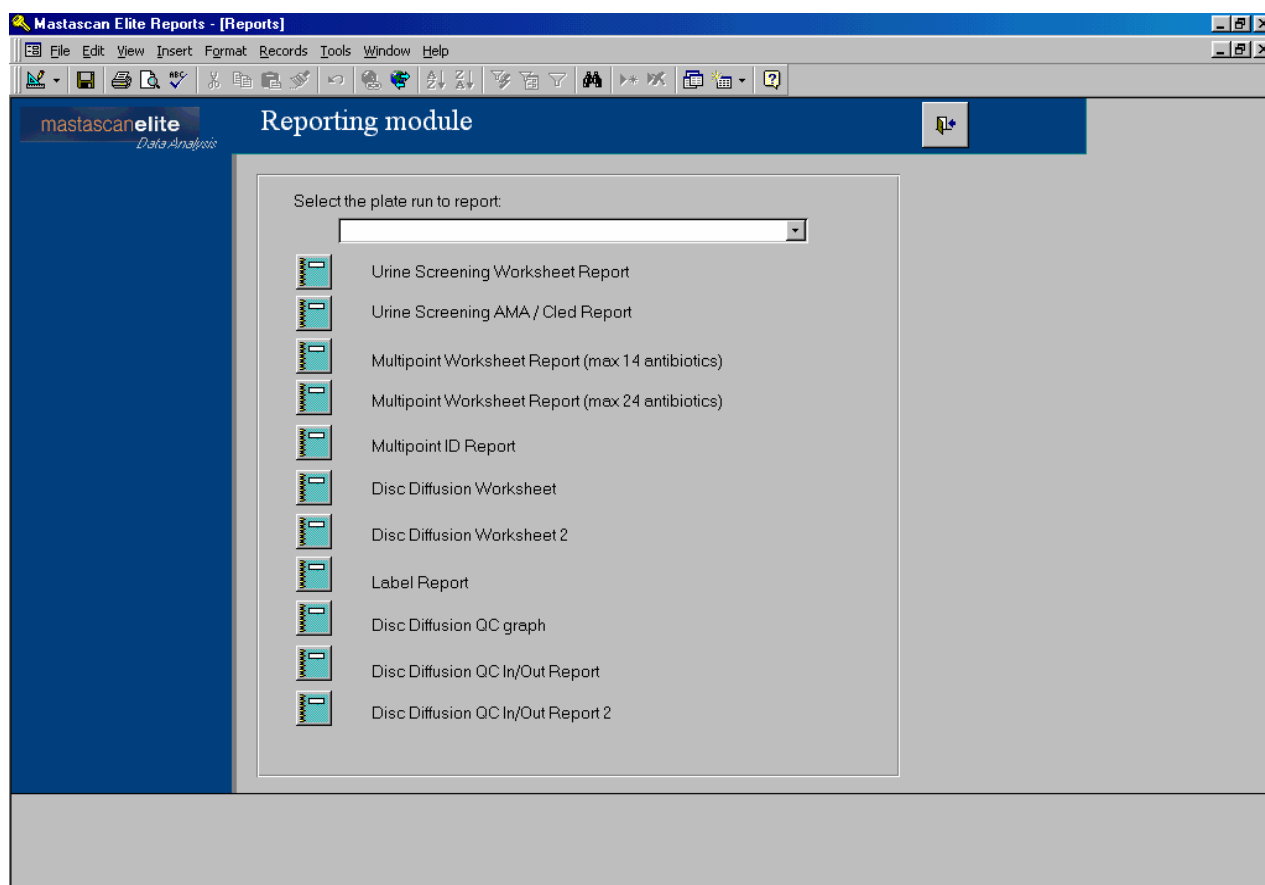
Once all the relevant plates have been moved to the right side of the window click on the **Export** button to send the data to the Laboratory Host System.

Once all the data has been exported click the **OK** button on the pop up message.

Close the main Import/Export window.

PRINTING THE RESULTS USING REPORTS

On the desktop double click on the icon labelled Reports, this will open the Reports



Database (figure5.3)

Figure 5.3. Reports Database

Select a plate run to be viewed from the drop down menu.

Then click on a relevant report button for Urine Screening they are:

- Urine Screening Worksheet Report
- Urine Screening AMA/Cled Report
- Multipoint Work Sheet (max14 or max 24)
- Multipoint ID Report

Although the reports are of a fixed layout they can be customised and this is done after the system is installed and calibrated.

Click on the Printer icon to print off the results for the selected plate run. Ensure a suitable Microsoft Windows operated printer is attached to the PC before printing.



Trouble Shooting

PROBLEMS AND SOLUTIONS

Problem	Solution
When viewing a plate on the machine the image is not centred.	Open the drawer and check to see if the plate has been aligned properly in the drawer Re-adjust the position of the plate
When I press any of the buttons on the light box, nothing happens.	Check the power switch on the back of the machine is on and the switch glows read. Check the electricity supply to the machine working. Check all external cables are connected properly Re-boot the PC ensuring the light box is already on.
The image is lighter at the on the left and right side of the plates	Check the side lighting blocker is in place on the drawer



Chapter 6

MAST GROUP LTD

For more advice and help there many ways to contact Mast Group.

TELEPHONE

IT HELPDESK **+44 (0)151 472 1473**

This line is open from 9.00 am to 5.00pm Monday to Friday

MAIN SWITCHBOARD **+44 (0)151 933 7277**

This line is open from 9.00 am to 5.00 PM Monday to Friday with an answer machine all other hours.

FAX

FAX **+44 (0) 151 472 1429**

POST

mastascanelite™ Product Support Service

Mast Group

Atlantic House

Derby Road

Bootle

Merseyside L20 1EA

United Kingdom

e-mail

E-MAIL **support@mastgrp.com**

If contacting by E-mail your correspondence will be dealt with within normal office ours on the same day or the next working day.



Important Information

The **mastascanelite™** is operated using electrical mains of 220-240VAC. Under no circumstances should the cover be removed from the **mastascanelite™** reading box. There is a serious risk of electrocution if removed.

Only a Service Engineer from Mast Group Ltd or an appropriately appointed Service Engineer working on behalf of Mast Group Ltd is authorised to open the instrument and perform any repairs, maintenance or cleaning

Any unauthorised opening of the **mastascanelite™** reading box will result in any existing warranties or maintenance contracts becoming null and void and the User will be liable for any risk to health due to potential electrical hazards. The cost of repair for any damage to the **mastascanelite™** due to customer intervention will be fully chargeable to the customer.

The **mastascanelite™** has no user servicable parts.

The **mastascanelite™** bears the CE mark indicating compliance with:

EMC Directive 89/336/EEC

The CE mark is not intended to indicate compliance with :

Directive 98/79/EEC In Vitro Diagnostic Medical Devices